

# Near-Complete Genome Sequences of Eight Human Astroviruses Recovered from Diarrheal Stool Samples of Hospitalized Children in Coastal Kenya in 2019

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**ABSTRACT** Here, using a sequence-independent sequencing approach (M. V. Phan, P. Hong Anh, N. Van Cuong, B. Oude Munnink, et al., *Virus Evol* 2:vew027, 2016, <https://doi.org/10.1093/ve/vew027>), we determined human astrovirus (HAstV) genome sequences from eight diarrheal stool samples collected in coastal Kenya in 2019. Phylogenetic analysis identified the following 4 genotypes: HAstV-1 ( $n = 4$ ), HAstV-2 ( $n = 1$ ), HAstV-3 ( $n = 1$ ), and HAstV-5 ( $n = 2$ ).

**H**uman astroviruses (HAstVs) (family *Astroviridae*) are nonenveloped, 7-kb positive-sense, single-stranded RNA genome viruses (1) and are among the top 5 viral causes of childhood diarrhea globally (2). HAstV clinical isolates are classified into classic HAstVs (HAstV-1 to HAstV-8), HAstV-MLB, and HAstV-VA/HMO (1).

In Kenya and other African settings, HAstV positivity in children with diarrhea as one of their illness symptoms ranges from 2.7% to 10.3% (3–5). To date, there are no complete or near-complete ( $\geq 90\%$  genome coverage) HAstV genome sequences from East Africa in the GenBank database (6). Analysis of HAstV genome sequences may facilitate optimization of molecular diagnostics and tracking the spread of HAstVs (7). Here, we utilized sequence-independent single-primer amplification (SISPA) sequencing to generate new HAstV genome sequences from positive reverse transcription-quantitative PCR (RT-PCR) (5) samples collected from children hospitalized with diarrhea in Kilifi, Kenya.

Total nucleic acid (TNA) was extracted from the 10 stool specimens using the QIAamp fast DNA stool minikit (Qiagen, Manchester, United Kingdom). The TNA was treated with Turbo DNase (Invitrogen, Carlsbad, CA), and first-strand synthesis was performed with FR26RV-ENDOH primers (8). Second-strand DNA synthesis was performed with Klenow fragment 3' to 5' exo- (New England BioLabs). To achieve a nonselective nucleic acid amplification, double-stranded DNA (dsDNA) was primed with the FR20RV primer (5'-GCCGAGCTCTGCAGATATC-3'), complementary to the FR26RV-ENDOH primers at the 5' end (9), and amplified using SuperScript III with the Platinum *Taq* DNA polymerase kit (Qiagen) as per the manufacturer's protocol. The PCR product was used to prepare Illumina barcoded libraries using the Illumina DNA Flex kit and sequenced in one run using the Illumina MiSeq machine generating 75-bp paired-end reads. Sequencing adapters and low-quality bases (Phred score,  $< 30$ ) were trimmed/removed from the short-read data using QUASR v.7.03 (10). Reference HAstV-1, HAstV-2, HAstV-3, and HAstV-5 genome sequences (GenBank accession numbers [JF327666](#), [KF039911](#), [MN444721](#), and [MF684776](#), respectively) were used for reference-guided assembly and to transfer annotations to the assembled genomes using the inbuilt Geneious mapper

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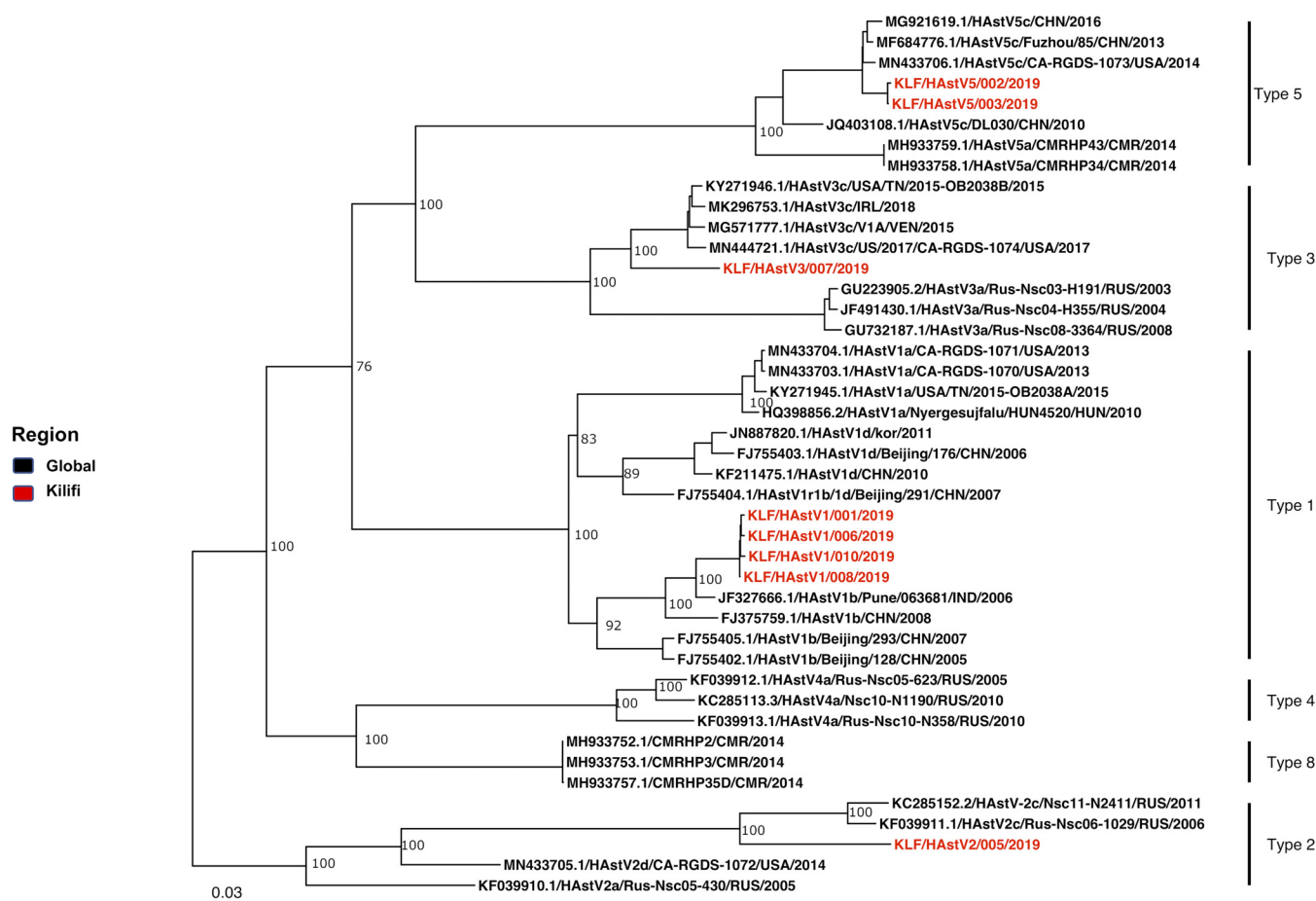
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**TABLE 1** Characteristics of human astrovirus genomes from coastal Kenya in 2019

Strain	Type	Collection date (day-mo-yr)	C <sub>T</sub> value <sup>a</sup>	Age (mo)	Sex	Symptom(s) <sup>b</sup>	Genome length (nt) <sup>c</sup>	Total no. of raw reads	No. of mapped reads	Avg depth <sup>d</sup>	Genome coverage (%) <sup>e</sup>	GC content (%)	Painwise identity to reference (%)	GenBank accession no.	Reference genome length (nt)
KLF/ASV/001	HAsV1	13/4/2019	19.5	25	Female	D + V	6,115	2,014,832	179,076	138	90.24	44.3	97.2	MW485038	6,776
KLF/ASV/008	HAsV1	26/4/2019	22.8	23	Male	D + V	6,776	1,297,222	5,673	53	100.00	44.9	97.4	MW485040	6,776
KLF/ASV/010	HAsV1	18/7/2019	22.5	15	Male	D + V	6,398	1,317,294	878	9	94.42	47.9	97.2	MW485041	6,776
KLF/ASV/006	HAsV1	23/7/2019	21.4	8	Male	D + V	6,698	3,015,006	3,744	39	98.85	45.0	97.3	MW485039	6,776
KLF/ASV/009	HAsV1	10/6/2019	24.0	27	Female	D + V	5,342	1,912,848	388	5	78.84				6,776
KLF/ASV/004	HAsV1	19/6/2019	22.2	10	Female	D + V	4,788	1,952,244	195	3	70.66				6,776
KLF/ASV/005	HAsV2	19/6/2019	23.9	12	Male	D + V	6,725	1,581,106	13,121	158	99.22	44.2	90.3	MW485042	6,778
KLF/ASV/007	HAsV3	15/4/2019	26.2	22	Female	D	6,747	2,302,640	10,137	100	99.37	44.0	94.1	MW485043	6,790
KLF/ASV/002	HAsV5	1/6/2019	24.3	9	Female	D	6,666	2,906,060	1,769	18	97.99	43.6	98.3	MW485044	6,803
KLF/ASV/003	HAsV5	1/6/2019	22.4	24	Male	D + V	6,361	3,046,380	1,188	13	93.50	43.7	98.5	MW485045	6,803

<sup>a</sup>The real-time RT-PCR (RPT-PCR) assay, including primers and probe sequences used for HAsV detection, has been described previously (6). C<sub>T</sub>, cycle threshold.<sup>b</sup>Objective evidence of a diarrheal disease. D, diarrhea; V, vomiting.<sup>c</sup>nt, nucleotide.<sup>d</sup>Calculated by dividing the per-position coverage output by respective genome length.<sup>e</sup>Calculated by dividing the genome length by the respective reference genome length.



**FIG 1** Maximum likelihood phylogenetic tree based on the open reading frame (ORF) sequences of the eight classical HAdVs (>90% genome coverage) identified in this study and representative strains from GenBank. The tree was constructed using IQ-Tree v.2.0.6 (13) with standard model selection. Bar indicates nucleotide substitutions per site. Red and black show HAdVs identified in this study and globally, respectively.

and annotation tools, respectively, on Geneious Prime v.2019.2.3 (11). MAFFT v.7.313 (12) was used for nucleotide coding sequence alignment, and maximum likelihood phylogenies were reconstituted in IQ-Tree v.2.0.6 (13) with standard model selection. Written informed consent for study participation was obtained from parents/guardians of the enrolled children, and the study protocol was approved by the KEMRI Scientific and Ethics Review Unit (SSC 2861 and SERU CGMRC/113/3624).

Patient demographics and sequencing output characteristics for the 10 samples are provided in Table 1. Eight samples yielded a consensus sequence covering >90% of the HAdV full-length genome. A maximum likelihood phylogeny of these eight near-complete genomes, including all publicly available HAdV genomes, is shown in Fig. 1. The new Kilifi sequences clustered with four different types of classical HAdVs, namely, HAdV-1 ( $n = 4$ ), HAdV-2 ( $n = 1$ ), HAdV-3 ( $n = 1$ ), and HAdV-5 ( $n = 2$ ). Both the HAdV-1 ( $n = 4$ ) and HAdV-5 ( $n = 2$ ) genomes had > 99% nucleotide similarity within their respective types. These new near-complete HAdV genomes from coastal Kenya increase available HAdV genomic data to support future molecular studies and local diagnostic methods.

**Data availability.** The raw sequence data were deposited in the Sequence Read Archive (SRA) under BioProject accession number [PRJNA692787](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA692787) and BioSample accession numbers [SAMN17370496](https://www.ncbi.nlm.nih.gov/biosample/SAMN17370496) to [SAMN17370503](https://www.ncbi.nlm.nih.gov/biosample/SAMN17370503). The genome sequences generated here were deposited in GenBank under accession numbers [MW485038](https://www.ncbi.nlm.nih.gov/nuccore/MW485038) to [MW485045](https://www.ncbi.nlm.nih.gov/nuccore/MW485045).

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## REFERENCES

- Donato C, Vijaykrishna D. 2017. The broad host range and genetic diversity of mammalian and avian astroviruses. *Viruses* 9:102. <https://doi.org/10.3390/v9050102>.
- Olortegui MP, Rouhani S, Yori PP, Salas MS, Trigoso DR, Mondal D, Bodhidatta L, Platts-Mills J, Samie A, Kabir F, Lima A, Babji S, Shrestha SK, Mason CJ, Kalam A, Bessong P, Ahmed T, Mduma E, Bhutta ZA, Lima I, Ramdass R, Moulton LH, Lang D, George A, Zaidi AKM, Kang G, Houpt ER, Kosek MN, on behalf of the MAL-ED Network. 2018. Astrovirus infection and diarrhea in 8 countries. *Pediatrics* 141:e20171326. <https://doi.org/10.1542/peds.2017-1326>.
- Lekana-Douki SE, Kombila-Koumavor C, Nkoghe D, Drosten C, Drexler JF, Leroy EM. 2015. Molecular epidemiology of enteric viruses and genotyping of rotavirus A, adenovirus and astrovirus among children under 5 years old in Gabon. *Int J Infect Dis* 34:90–95. <https://doi.org/10.1016/j.ijid.2015.03.009>.
- Nguekeng Tsague B, Mikounou Louya V, Ntouni F, Adedjoja A, Vouvongui CJ, Peko SM, Abena AA. 2020. Occurrence of human astrovirus associated with gastroenteritis among Congolese children in Brazzaville, Republic of Congo. *Int J Infect Dis* 95:142–147. <https://doi.org/10.1016/j.ijid.2020.02.056>.
- Lambisia AW, Onchaga S, Murunga N, Lewa CS, Nyanjom SG, Agoti CN. 2020. Epidemiological trends of five common diarrhea-associated enteric viruses pre-and post-rotavirus vaccine introduction in coastal Kenya. *Pathogens* 9:660. <https://doi.org/10.3390/pathogens9080660>.
- Federhen S. 2012. The NCBI taxonomy database. *Nucleic Acids Res* 40: D136–D143. <https://doi.org/10.1093/nar/gkr1178>.
- Yinda CK, Shi C, Deboutte W, Conceição-Neto N, Van Ranst M, Beller L, Maes P, Vanhulle E, Ghogomu SM, Matthijnsens J. 2019. Gut virome analysis of Cameroonians reveals high diversity of enteric viruses, including potential interspecies transmitted viruses. *mSphere* 4:e00585-18. <https://doi.org/10.1128/mSphere.00585-18>.
- Nguyen AT, Tran TT, Hoang VMT, Nghiem NM, Le NNT, Le TTM, Phan QT, Truong KH, Le NNT, Ho VL, Do VC, Ha TM, Nguyen HT, Nguyen CVV, Thwaites G, Van Doorn HR, Van Le T. 2016. Development and evaluation of a non-ribosomal random PCR and next-generation sequencing based assay for detection and sequencing of hand, foot and mouth disease pathogens. *Virol J* 13:125. <https://doi.org/10.1186/s12985-016-0580-9>.
- Djikeng A, Halpin R, Kuzmickas R, DePasse J, Feldblyum J, Sengamalay N, Afonso C, Zhang X, Anderson NG, Ghedin E, Spiro DJ. 2008. Viral genome sequencing by random priming methods. *BMC Genomics* 9:5. <https://doi.org/10.1186/1471-2164-9-5>.
- Watson SJ, Welkers MRA, Depledge DP, Coulter E, Breuer JM, de Jong MD, Kellam P. 2013. Viral population analysis and minority-variant detection using short read next-generation sequencing. *Philos Trans R Soc B Biol Sci* 368:20120205. <https://doi.org/10.1098/rstb.2012.0205>.
- Biomatters Limited. 2018. Geneious—molecular biology and NGS analysis tools. Biomatters Limited, Auckland, New Zealand.
- Katoh K, Standley DM. 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Mol Biol Evol* 30:772–780. <https://doi.org/10.1093/molbev/mst010>.
- Nguyen LT, Schmidt HA, Von Haeseler A, Minh BQ. 2015. IQ-TREE: a fast and effective stochastic algorithm for estimating maximum-likelihood phylogenies. *Mol Biol Evol* 32:268–274. <https://doi.org/10.1093/molbev/msu300>.